

CE 421/521 Environmental Biotechnology - Fall 2008

Biogeochemical Cycles (C, N, P, & S)

- composition of bacterial cell (molar formula: $C_5H_7O_2N$ with P 1/5 of the N requirement)
- limiting nutrients are _____ and _____

TABLE 14.1 Chemical Composition of an *E. coli* Cell

Elemental breakdown	% dry mass of an <i>E. coli</i> cell
Major elements	
Carbon	50
Oxygen	20
Hydrogen	8
Nitrogen	14
Sulfur	1
Phosphorus	3
Minor elements	
Potassium	2
Calcium	0.05
Magnesium	0.05
Chlorine	0.05
Iron	0.2
Trace elements	
Manganese	all trace elements combined comprise 0.3% of dry weight of cell
Molybdenum	
Cobalt	
Copper	
Zinc	

Adapted from Neidhardt *et al.* (1990).

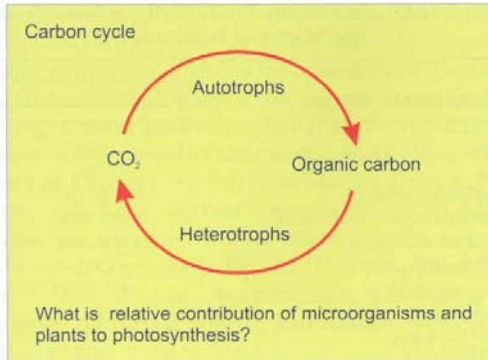


FIGURE 14.1 The carbon cycle is dependent on autotrophic organisms that fix carbon dioxide into organic carbon and heterotrophic organisms that respire organic carbon to carbon dioxide.

TABLE 14.3 Global Carbon Reservoirs

Carbon reservoir	Metric tons carbon	Actively cycled
Atmosphere		
CO ₂	6.7×10^{11}	Yes
Ocean		
Biomass	4.0×10^9	No
Carbonates	3.8×10^{13}	No
Dissolved and particulate organics	2.1×10^{12}	Yes
Land		
Biota	5.0×10^{11}	Yes
Humus	1.2×10^{12}	Yes
Fossil fuel	1.0×10^{13}	Yes
Earth's crust ^a	1.2×10^{17}	No

^a This reservoir includes the entire lithosphere found in either terrestrial or ocean environments. (Data from Dobrovolsky, 1994.)

TABLE 14.4 Net Carbon Flux between Selected Carbon Reservoirs

Carbon source	Flux (metric tons carbon/year)
Release by fossil fuel combustion	7×10^9
Land clearing	3×10^9
Forest harvest and decay	6×10^9
Forest regrowth	-4×10^9
Net uptake by oceans (diffusion)	-3×10^9
Annual flux	9×10^9

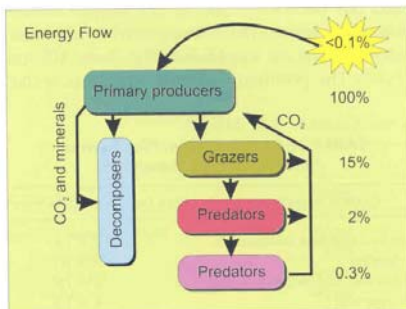


FIGURE 14.2 Diagram of the efficiency of sunlight energy flow from primary producers to consumers.

Table 14.6 Major Types of Organic Components of Plants

Plant component	% dry mass of plant
Cellulose	15–60
Hemicellulose	10–30
Lignin	5–30
Protein and nucleic acids	2–15

NITROGEN

- Atmosphere is _____% nitrogen, yet nitrogen is considered a l_____ n_____
- required in p_____

TABLE 14.10 Global Nitrogen Reservoirs

Nitrogen reservoir	Metric tons nitrogen	Actively cycled
Atmosphere		
N ₂	3.9 × 10 ¹⁵	No
Ocean		
Biomass	5.2 × 10 ⁹	Yes
Dissolved and particulate organics	3.0 × 10 ¹¹	Yes
Soluble salts (NO ₃ ⁻ , NO ₂ ⁻ , NH ₄ ⁺)	6.9 × 10 ¹¹	Yes
Dissolved N ₂	2.0 × 10 ¹³	No
Land		
Biota	2.5 × 10 ¹⁰	Yes
Organic matter	1.1 × 10 ¹¹	Slow
Earth's crust ^a	7.7 × 10 ¹⁴	No

^a This reservoir includes the entire lithosphere found in either terrestrial or ocean environments. (Adapted from Dobrovolsky, 1994.)

Fixation

- _____ metric tons/y compared to 2.5x10¹⁰ metric tons C/y
- c_____ & few others
 - non-s_____ – *Clostridia*
 - symbiotic – *Rhizobium*
- n_____ – requires Mg²⁺ & ATP (15 to 20 ATP/N₂)

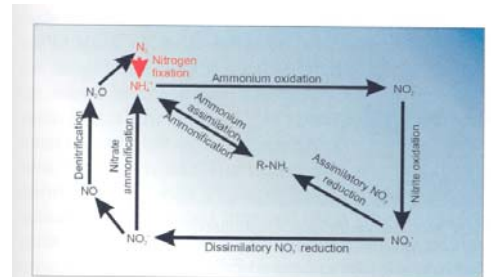
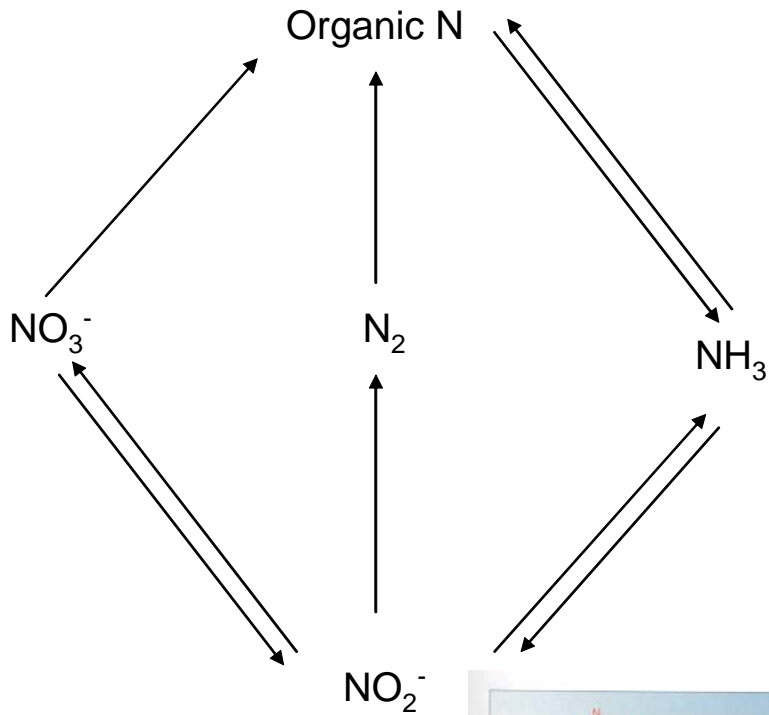
Assimilation – NH₃ (NH₄⁺) preferred, will use NO₃⁻ but has to be reduced to NH₄⁺

- C/N ratio is approximately _____:1 for aerobes
- C/N ratio is approximately _____:1 for anaerobes
- C/N ratio is approximately _____:1 for anaerobes in highly loaded (high rate) system
- cell c_____ is characterized by the empirical formula: C H O N with the P requirement as _____ the N requirement (alternatively C₆₀H₈₇O₂₃N₁₂P)
- in general cell composition is

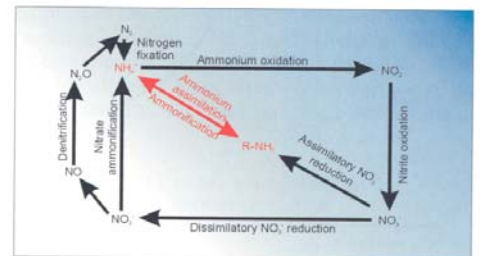
- * 50%
- * 20%
- * 10-15%
- * 8-10%
- * 1-3%
- * 0.5-1.5%

Ammonification: breakdown of o_____ N to inorganic nitrogen

ex. urea:



Summary:
 N₂ fixation is energy intensive
 End product of N₂ fixation is ammonia
 N₂ fixation is inhibited by ammonia
 Nitrogenase is O₂ sensitive, some free-living N₂ fixers require reduced O₂ tension



Summary:
 Assimilation and ammonification cycles ammonia between its organic and inorganic forms
 Assimilation predominates at C:N ratios > 20
 ammonification predominates at C:N ratios < 20

parameter	Nitrosomonas		Nitrobacter	
	range	typical (@ 20°C)	range	typical (@ 20°C)
μ_{max}	0.014 - 0.092	0.032	0.006 - 0.06	0.034
K_S	0.06 - 5.6	1.0	0.06 - 8.4	1.3
K_O	0.3 - 1.3	0.5	0.3 - 1.3	0.68
Yield	0.04 - 0.13	0.1	0.02 - 0.07	0.05

Optimum pH for nitrifiers is around 8.0, range 7.5 - 8.5 (higher than for most other biological processes).

Nitrifiers are sensitive to

- d _____ o _____
- t _____
- p _____
- i _____

$$\mu = \frac{\mu_{max} S_{NH4}}{K_S + S_{NH4}} \cdot \frac{K_I}{K_I + I}$$

where I = concentration of inhibitor, mg/L
 K_I = inhibition coefficient, mg/L

Effects of Temperature

derivation of the Arrhenius equation $k = Ae^{\frac{-\mu}{RT}}$

$$k_2 = k_1 \theta^{(T_2 - T_1)}$$

where $k_{1,2}$ = reaction rate coefficient at temperature $T_{1,2}$

$\theta = \frac{k_2}{k_1} = e^{\frac{-\mu}{R} \left(\frac{1}{T_2} - \frac{1}{T_1} \right)}$

theta values

	Nitrosomonas	Nitrobacter
μ_{max}	1.098 - 1.118	1.068 - 1.112
K_S	1.125	1.157
k_d	1.029 - 1.104	1.029 - 1.104

given the following measured data, calculate the theta value

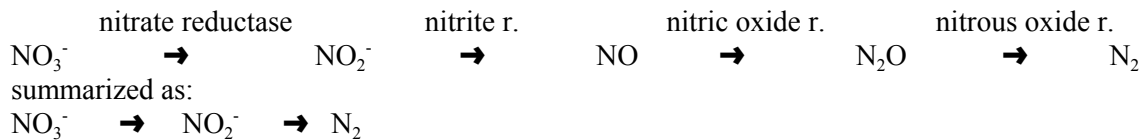
T, °C	b, h ⁻¹
10	0.0037
20	0.0095
30	0.0229
40	0.0372

DENITRIFICATION

1. A _____ nitrate reduction: $\text{NO}_3^- \rightarrow \text{NH}_4^+$ nitrate is incorporated into cell material and reduced inside the cell

2. D _____ nitrate reduction (denitrification)

NO_3^- serves as the t _____ e _____ a _____ (TEA) in an anoxic (anaerobic) environment



requires o _____ m _____ (example: methanol)

$$\mu = \frac{\mu_{\max} S}{K_S + S} \cdot \frac{\text{NO}_3^-}{K_{\text{NO}_3} + \text{NO}_3^-} \quad \text{calculate COD of methanol:} \quad 6 \text{NO}_3^- + 5\text{CH}_3\text{OH} \rightarrow 3\text{N}_2 + 5\text{CO}_2 + 7\text{H}_2\text{O} + 6\text{OH}^-$$

calculate alkalinity:

kinetics for denitrification similar to those for heterotrophic aerobic growth

Nitrogen Removal in Wastewater Treatment Plants

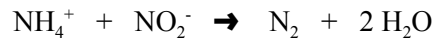
Total Kjeldahl Nitrogen (TKN) = o_____ n_____ + a_____
(measured by digesting sample with sulfuric acid to convert all nitrogen to ammonia)

- TKN ~ 35 mg/L in influent
- p_____ t_____ removes approximately 15%
- additional removal with biomass w_____

3 Methods for Nitrogen Removal

1. Biological

- n_____
- d_____
- ANAMMOX: ammonium is the electron donor, nitrite is the TEA



2. Chemical/Physical

- air s_____
- breakpoint c_____
- ion e_____
- reverse o_____

Concerns for nitrogen discharge:

1. T_____
2. D_____ of DO
3. E_____
4. Nitrate in d_____ water – causes methemoglobinemia (blue baby) oxidizes hemoglobin to methemoglobin

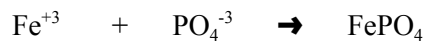
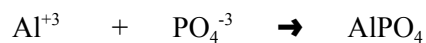
PHOSPHORUS

- limiting n_____ in algae (at approximately 1/5 the nitrogen requirement)
- 15% of population in US discharges to l_____
- wastewater discharge contains approximately 7- 10 mg/L as P
- o_____
- i_____: orthophosphate

Removal of Phosphorus

1. Chemical precipitation:

- a. traditional p_____ reactions



- b. as s_____ (magnesium ammonium phosphate, MAP)



2. Enhanced Biological Phosphorus Removal (EBPR) see handout

SULFUR

— inorganic: SO_4^{-2} S° H_2S

— organic: $\text{R} - \text{O} - \text{SO}_3^{-2}$

— four key reactions:

1. H_2S o _____ — can occur aerobically or anaerobically to elemental sulfur (S°)

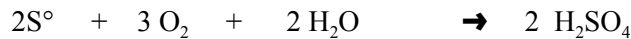
a. a _____: *Thiobacillus thioparus* oxidizes S^{-2} to S°



b. a _____: — phototrophs use H_2S as electron donor

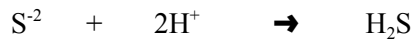
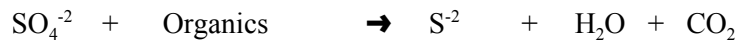
S° in sulfur — filamentous sulfur bacteria oxidize H_2S to granules: *Beggiatoa*, *Thiothrix*

2. Oxidation of E _____ Sulfur (*Thiobacillus thiooxidans* at low pH)



3. A _____ sulfate reduction: proteolytic bacteria breakdown organic matter containing sulfur (e.g. amino acids: methionine, cysteine, cystine)

4. D _____ sulfate reduction: under anaerobic conditions — s _____ r _____ b _____ (SRB)



— *Desulvibrio* and others

— Sulfate is used as a TEA & l _____ m _____ w _____ organics serve as the electron donors

TABLE 14.15 Genera of Denitrifying Bacteria

Genus	Interesting characteristics
Organotrophs	
<i>Alcaligenes</i>	Common soil bacterium
<i>Agrobacterium</i>	Some species are plant pathogens
<i>Aquaspirillum</i>	Some are magnetotactic, oligotrophic
<i>Azospirillum</i>	Associative N_2 fixer, fermentative
<i>Bacillus</i>	Spore former, fermentative, some species thermophilic
<i>Blastobacter</i>	Budding bacterium, phylogenetically related to <i>Rhizobium</i>
<i>Bradyrhizobium</i>	Symbiotic N_2 fixer with legumes
<i>Branhamella</i>	Animal pathogen
<i>Chromobacterium</i>	Purple pigmentation
<i>Cytophaga</i>	Gliding bacterium; cellulose degrader
<i>Flavobacterium</i>	Common soil bacterium
<i>Flexibacter</i>	Gliding bacterium
<i>Halobacterium</i>	Halophilic
<i>Hyphomicrobium</i>	Grows on one-C substrates, oligotrophic
<i>Kinoshella</i>	Animal pathogen
<i>Neisseria</i>	Animal pathogen
<i>Paracoccus</i>	Halophilic, also lithotrophic
<i>Propionibacterium</i>	Fermentative
<i>Pseudomonas</i>	Commonly isolated from soil, very diverse genus
<i>Rhizobium</i>	Symbiotic N_2 fixer with legumes
<i>Wolfinella</i>	Animal pathogen
Phototrophs	
<i>Rhodospseudomonas</i>	Anaerobic, sulfate reducer
Lithotrophs	
<i>Alcaligenes</i>	Uses H_2 , also heterotrophic, common soil isolate
<i>Bradyrhizobium</i>	Uses H_2 , also heterotrophic, symbiotic N_2 fixer with legumes
<i>Nitrosomonas</i>	NH_3 oxidizer
<i>Paracoccus</i>	Uses H_2 , also heterotrophic, halophilic
<i>Pseudomonas</i>	Uses H_2 , also heterotrophic, common soil isolate
<i>Thiobacillus</i>	S-oxidizer
<i>Thiomicrospira</i>	S-oxidizer
<i>Thiosphaera</i>	S-oxidizer, heterotrophic nitrifier, aerobic denitrification

From Myrold, 1998.

— Low cell y _____

— P _____ of SRB depends on COD:S ratio, particularly readily degradable (e.g., VFA) COD

— SRB compete with m _____ for substrate: high COD:S favors methanogens, low COD:S favors SRB

Crown Sewer Corrosion

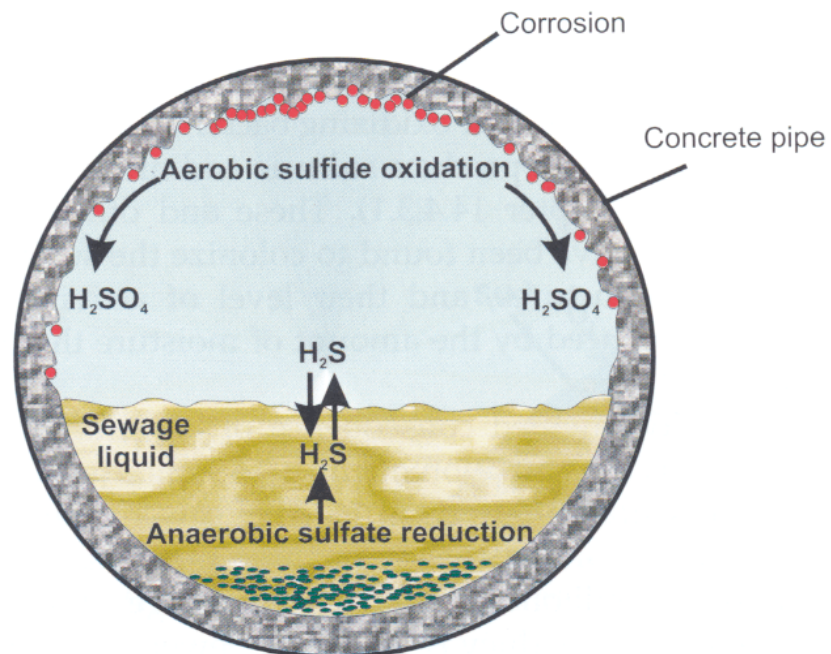


FIGURE 15.3 Cross section showing microbial involvement in the corrosion of a concrete sewer pipe. (Adapted from Sydney *et al.*, 1996.)